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INVESTIGATION OF THE COAGULATION AND ADHESION PHENOMENA IN COLLOIDS BY FIELD-FLOW FRACTIONATION

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ABSTRACT

Conventional and potential barrier methodologies of Field-Flow Fractionation are used to separate and characterize colloidal particles of haematite and titanium dioxide.

The colloidal particles are reversibly adsorbed at the beginning of the Sedimentation Field-Flow Fractionation channel wall. The reversibility of the adsorption of these particles can be achieved by the appropriate adjustment of various parameters among which the must suitable is the ionic strength of the carrier solution.

In the present work by using both Field-Flow Fractionation (FFF) and Transmission Electron Microscopy (TEM) techniques we investigate whether the reversible adsorption of the haematite and titanium dioxide particles on the stainless-steel channel wall is due to their reversible coagulation or to the variation of the interaction forces between the colloidal particles and the solid surface.

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INTRODUCTION

In Field-Flow Fractionation (FFF) components are separated due to their differential interaction with an applied field.¹⁴ Sedimentation Field-Flow Fractionation (SdFFF) is capable of separating and characterizing colloidal and particulate materials.^{5, 6} It is a relatively new technique for the high resolution separation and analysis of a wide variety of colloids and macromolecules.³⁻⁶

Samples are separated in a thin channel on the basis of their effective masses. The flow through the channel has a parabolic profile whose flow velocity approaches zero at both walls and reaches a maximum at the midpoint between the walls. When a centrifugal force is applied across the channel, particles are driven toward the bottom wall, and an equilibrium distribution is established between the external force and diffusion of the particles.⁵ Since the equilibrium distribution will determine the average velocity for a sample zone eluted through the channel, the sample particles elute in order of decreasing diffusion coefficient, i.e. the smaller particles elute first. When the average distance from the accumulation wall is exceeded by the particle radius, the steric-hyperlayer effect has to be considered and the particle velocity will increase with increasing diameter. As a consequence, the onset of the steric mechanism causes a shift in the elution order, so that the larger particles elute first.^{7,9}

Potential-barrier Field-Flow Fractionation (PBFFF) is a new method,^{10,11} which is based either on particle size differences or on Hamaker constant, surface potential^{12,14} and Debye-Huckel reciprocal distance differences. Experimentally, this should be done by changing the ionic strength of the carrier solution^{10,11} from a high value, in which the colloidal particles are adsorbed at the beginning of the SdFFF channel wall, to a lower value, where the total number of adhered particles is released.^{10,11,15,16}

The PBFFF technique can be used successfully to separate and characterize dilute and concentrated colloidal samples even with the same size but with different limiting electrolyte concentrations for their adsorption on solid surfaces.

In the present work we investigated whether the reversible adsorption of the colloidal particles on the stainless-steel channel wall is due to their reversible coagulation or to the interaction forces between the colloids and the solid surface.

EXPERIMENTAL

The basic sedimentation FFF system used for these studies has been described in detail previously.^{13, 17, 18} Three channels having different dimensions

were used in the present work. The first (channel-1) had a length of 38.4 cm, breadth of 2.05 cm, and thickness of 0.0262 cm. The void volume of the channel-1 measured by the elution of non-retained peak of sodium benzoate was found to be 2.06 cm³. The second channel (channel-2) had the following dimensions: $38.5 \times 2.3 \times 0.0181$ cm³ with a channel void volume of 1.60 cm³. The third channel (channel-3) had breadth of 2.35 cm, thickness of 0.0193 cm and the same length as the channel-1. The void volume, measured by the same method, was 1.74 cm³.

In all systems the column was 6.85 cm from the centre of rotation. A Gilson Holochrome UV detector was used for detection at 254 nm and a Gilson Minipuls 2 peristaltic pump was used to pump the carrier solution and the sample to the channels.

Titanium dioxide monodisperse colloidal particles from PolySciences with an average diameter of d = 0.310 μ m (obtained by TEM), which were studied in channel-1, and haematite nearly monodisperse colloidal particles of two sizes provided by Prof. J. Lyklema (Agricultural University, Wageningen, The Netherlands) with an average diameter (obtained by TEM) of d = 0.146 μ m [a-Fe₂O₃(I)] and d = 0.258 μ m [a-Fe₂O₃(II)], which were studied in channels 2 and 3, respectively, were used as samples. The morphology of the sample particles were investigated by Phillips CM-20 transmission electron microscopy (TEM) at 200 kV.

The carrier solution was triply distilled water containing 0.5% (v/v) of detergent FL-70 (Fisher Scientific Co.) and 0.02% (w/w) sodium azide as bacteriocide. Potassium nitrate from Riedel-De Haen A.G., was added to the carrier solution for the adjustment of the ionic strength.

The experimental procedure was described in detail elsewhere.¹¹ The critical electrolyte (KNO₃) concentration of the carrier solution for the total adsorption of TiO₂ and a-Fe₂O₃ (II) on the stainless-steel channel was found to be 3×10^{-2} M, while that for the adsorption of a-Fe₂O₃(I) particles was found to be 8×10^{-2} M.

RESULTS AND DISCUSSION

The basic retention relationship in FFF shows that the retention ratio *R* is a function only of the dimensionless retention parameter $\lambda^{3, 19, 20}$:

$$R = 6\lambda \left[\coth(1/2\lambda) - 2\lambda \right] = \frac{V_o}{V_r}$$
(1)



Figure 1. Fractograms of monodisperse TiO_2 spherical particles obtained (a) by conventional sedimentation FFF and (b) by potential barrier sedimentation FFF.



Figure 2. Transmission electron micrographs (TEMs) of TiO₂ particles (20000x) dispersed (a) in 0.5% v/v FL-70 + 1×10^3 M KNO₃ and (b) in 0.5% v/v FL-70 + 3×10^2 M KNO₃.

where V_{o} is the channel void volume and V_{r} is the retention volume of retained particles.

When $\lambda \ll 1$, Equation (1) reduces to the simple limiting form:

$$\lim_{\lambda \to 0} \mathbf{R} \approx 6\lambda \tag{2}$$

In conventional Sedimentation FFF (SdFFF), where the interactions between the colloidal particles and the channel wall are absent, any observed

value of V_r or R can be converted into the diameter d of a spherical particle via the equation:

$$d = \left(\frac{36kTV_r}{\pi Gw\Delta\rho V_o}\right)^{1/3}$$
(3)

where k is Boltzmann's constant, T is absolute temperature, G is the field strength expressed as acceleration, w is the channel thickness and $\Delta \rho$ is the density difference between the particles and the carrier solution.

In Potential Barrier FFF (PBFFF) by changing the total potential energy of interaction between the colloidal particles and the channel wall one can succeed the adhesion of the whole number of colloidal particles on the solid surface, which can be followed by the release of the adhered particles. Experimentally, this can be done by changing the surface potential of the particles, Ψ_1 , or the Hamaker constant, A, and the Debye-Huckel reciprocal distance, $\kappa^{-1} = BI^{1/2}$, as the total potential energy, *U*, of a spherical particle is given by the expression:¹¹

$$U = \frac{4}{3}\pi r^{3}\Delta Gx + \frac{A}{6} \left[ln \left(\frac{H+2r}{H} \right) - \frac{2r(H+r)}{H(H+2r)} \right] + 16\epsilon r \left(\frac{kT}{e} \right)^{2} tan h \left(\frac{e_{1}\psi_{1}}{4kT} \right) tan h \left(\frac{e\psi_{2}}{4kT} \right) e^{\frac{H}{Bl^{1/2}}}$$
(4)

where *r* is the particle radius, *x* is the coordinate position of the centre of particle mass, *H* is the separation distance between the sphere and the channel wall, *M* is the dielectric constant of the liquid phase, *e* is the electronic charge, Ψ_2 is the surface potential of the solid surface, *B* is a constant and *I* is the ionic strength of the carrier solution. In its simplest form the PBFFF technique consists in changing the ionic strength of the carrier solution from a high value, where the attraction between the colloidal particles and the channel leads to the total adsorption of the particles at the beginning of the SdFFF channel wall, to a lower value, where the total number of adhered particles is released.

In the present work the PBFFF technique was applied to the monodisperse spherical particles of titanium dioxide, as well as to the nearly monodisperse spherical particles of haematite. Figure 1a shows a fractogram obtained by the conventional SdFFF technique for the TiO₂ sample suspended in a carrier solution containing a low concentration of potassium nitrate (ca. 1×10^{-3} M). The retention volume was found to be 49.1 cm³ and calculated with the aid of

Table 1

Particle Diameters for the Sample of TiO₂, a-Fe₂O₃(I) and a-Fe₂O₃(II) Obtained by Transmission Electron Microscope at Various Dispersing Solutions

	Particle Diameter (µm)		
Dispersing Solution	TiO ₂	$a-Fe_2O_3(I)$	a-Fe ₂ O ₃ (II)
0.5% v/v FL-70		0.145	0.258
0.5% v/v FL-70 +	0.310	0.146	0.258
1 x 10 ⁻³ M KNO ₃			
0.5% v/v FL-70 +	0.309		0.248
$3 \times 10^{-2} M KNO_3$			
0.5% v/v FL-70 +		0.142	
8 x 10 ⁻² M KNO ₃			

Equation (3) diameter was $0.310 \ \mu\text{m}$. Changing the carrier solution to the one containing higher concentration of KNO₃ (ca. 3×10^{-2} M) led to the adsorption of the whole particle of TiO₂ on the channel wall. Finally, changing again the concentration of the carrier solution to the original one (ca. 1×10^{-3} M) led to the appearance of a sample peak, due to the desorption of the adhered particles (cf. Figure 1b).

The retention volume of this sample peak was 48.4 cm³ and the calculated particle diameter was found to be 0.309 μ m, very close to that found (0.310 μ m) from the direct injection of TiO₂ particles into the channel by the conventional SdFFF technique. The release of all the adhered TiO₂ particles during the PBFFF procedure was verified by the fact that no elution peak was obtained when the field strength was reduced to zero.

In order to show whether the adhesion of the TiO₂ particles on the channel wall is due to the their reversible coagulation, during the variation of the carrier solution ionic strength, TEM pictures for the TiO₂ sample dispersed in two different solutions were received (cf. Figure 2). The first solution contained detergent FL-70 (0.5% v/v) with 1×10^3 M KNO₃, in which no adsorption of the particles was observed, while the second detergent FL-70 (0.5% v/v) with 3×10^2 M KNO₃, in which the total number of TiO₂ particles is adhered at the beginning of the channel wall. The TEM pictures show that the TiO₂ particles have the same shape and size in both solutions ($0.310 \ \mu m$ in the carrier solution with 1×10^3 M KNO₃, cf. Table 1).



These values, which were measured from about 20 spheres in the TEM pictures, are exactly the same with those found by the PBFFF technique (0.309 μ m) or the conventional SdFFF technique (0.310 μ m) by the direct injection of TiO₂ particles into the channel, indicating that no reversible aggregation takes place during the variation of the ionic strength of the suspending medium. Thus, the total adhesion of the TiO₂ particles on the channel wall at concentrations of KNO₃ higher than 3×10^2 M can be attributed to the strong energy of attraction between the colloidal particles and the solid surface, which increases with the electrolyte concentration [cf. Equation (4)], and not to the coagulation of particles. It is obvious from Equation (3) that the increase of particle diameter due to the aggregation process leads to a higher increase in the retention volume. Thus the particles are moved along the column with a such low velocity, so that one could approximately consider that the particles are adhered at the beginning of the SdFFF channel wall.

The same phenomenon was investigated by using two samples of haematite [a-Fe₂O₃(I) with d = 0.146 μ m and a-Fe₂O₃(II) with d = 0.258 μ m] with nearly monodisperse spherical particles. Figure 3a shows a fractogram for the a-Fe₂O₃(I) sample obtained by the conventional SdFFF technique at a low electrolyte concentration (ca. 1 × 10⁻³ M KNO₃), in which no adsorption of the colloidal particles takes place at the beginning of the channel wall. The retention volume was found to be 24.8 cm³, which corresponds to a particle diameter of 0.146 μ m.

Changing the carrier solution to one containing a higher concentration of KNO₃ (ca. 8×10^{-2} M) led to the adsorption of all the a-Fe₂O₃(I) particles on the stainless-steel channel wall, as no elution curve was observed. When the carrier solution was changed to one containing a lower ionic strength (ca. 1×10^{-3} M KNO₃), a sample peak appeared as a consequence of the desorption of the a-Fe₂O₃(I) particles (cf. Figure 3b). The mean diameter of the a-Fe₂O₃(I) particles (0.148 µm) is very close to that obtained by the conventional SdFFF (0.146 µm), in which no adsorption of the colloidal particles takes place at the beginning of the channel wall.

The investigation of the adhesion process of the a-Fe₂O₃(I) particles on the SdFFF channel wall was done again by using the TEM technique. TEM pictures of the a-Fe₂O₃(I) particles dispersed in three different solutions (ca. 0.5% v/v FL-70, 0.5% v/v FL-70 + 1 × 10³ M KNO₃ and 0.5% v/v FL-70 + 8 × 10² M KNO₃) were received (cf. Figure 4).

Figure 3 (left). Fractograms of a-Fe₂O₃(I) colloidal particles obtained (a) by conventional sedimentation FFF and (b) by potential barrier sedimentation FFF.



Figure 4. Transmission electron micrographs (TEMs) of a-Fe₂O₃(I) particles (50000 x) dispersed (a) in 0.5% v/v FL-70 (b) in 0.5% v/v FL-70 + 10⁻³ M KNO₃ and (c) in 0.5% v/v FL-70 + 8 × 10⁻² M KNO₃.

The resulting mean diameters by measuring the size of about twenty (20) spheres in the TEM pictures, which are given in Table 1, are very close to each other, as well as to those determined by the conventional SdFFF (0.146 μ m) and the PBFFF (0.148 μ m) technique. The latter indicates that no aggregation of the



Figure 5. Fractograms of a-Fe $_2O_3(II)$ colloidal particles obtained (a) by conventional sedimentation FFF and (b) by potential barrier sedimentation FFF.



(a)



(c)

Figure 6. Transmission electron micrographs (TEMs) of a-Fe₂O₃(II) particles (50000 x) dispersed (a) in 0.5% v/v FL-70 (b) in 0.5% v/v FL-70 + 10^{-3} M KNO₃ and (c) in 0.5% v/v FL-70 + 3×10^{-2} M KNO₃.

 $a-Fe_2O_3(I)$ particles occurs at the dispersing medium with the higher electrolyte concentration (8 × 10⁻² M KNO₃) in which adsorption of the whole number of haematite particles takes place at the beginning of the SdFFF channel wall. Thus, the adhesion of a-Fe₂O₃(I) particles on the solid surface is due, as it was

also explained for the TiO_2 particles, to the increase of the van der Waals attraction potential, because of the increase of the ionic strength of the dispersing medium [cf. Equation (4)].

Finally, in order to show whether the particle size influences the total interaction potential between a spherical particle and a solid surface, as Equation (4) shows, we used a second sample of the same haematite material, $a-Fe_2O_3(II)$, but with different particle diameter $d = 0.258 \ \mu\text{m}$. Following the procedure described in the analysis of the previous two samples [TiO₂ and $a-Fe_2O_3(I)$], we found for the $a-Fe_2O_3(II)$ sample a particle diameter of 0.245 $\ \mu\text{m}$ by the conventional SdFFF technique, exactly the same as that determined by the PBFFF technique (cf. Figure 5). The critical electrolyte concentration at which adsorption of all of the $a-Fe_2O_3(II)$ colloidal particles occurs, at the beginning of the column was found to be 3×10^{-2} M KNO₃, [significantly lower from the value 8×10^{-2} M KNO₃ found for the adhesion of $a-Fe_2O_3(I)$ particles], while that at which the total number of adherent $a-Fe_2O_3(II)$ particles is released was 1×10^{-3} M KNO₃.

The measured mean particle diameters of the a-Fe₂O₃(II) sample at three different carrier solutions, (from the TEM pictures, cf. Figure 6) which are given in Table 1, are very close to each other (0.258 μ m for the carrier solution (c.s.) with only 0.5% v/v FL-70, 0.258 μ m for the c.s. with 0.5% v/v FL-70 + 1 $\times 10^3$ M KNO₃ and 0.248 μ m for the c.s. with 0.5% v/v FL-70 + 3 $\times 10^{-2}$ M KNO₃), as well as to those determined by the conventional SdFFF (0.245 μ m) and PBFFF (0.245 μ m) techniques. The latter indicates that during adhesion no aggregation phenomena of a-Fe₂O₃(II) particles take place and their total adsorption on the solid surface at high electrolyte concentrations can be attributed to the increase of the van der Waals attraction potential between the a-Fe₂O₃(II) particles and the sedimentation FFF channel wall.

In conclusion, although the TEM process suffers from the disadvantage of destroying the particle environment, the combination of the Field-Flow Fractionation technique with the Transmission Electron Microscopy method gives invaluable information concerning the coagulation and adhesion phenomena of colloidal materials.

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REFERENCES

- 1. J. C. Giddings, Science, 260, 1456 (1993).
- 2. K.D. Caldwell, Anal. Chem., 60, 959A (1988).
- 3. A. Koliadima, E. Dalas, G. Karaiskakis, J. High Resol. Chromatogr., **13**, 338 (1990).
- 4. J. J.Kirkland, W. W. Yan, W. A. Doerner, J. W. Grand, Anal. Chem., **52**, 1944 (1980).
- 5. J. C. Giddings, F. J. F. Yang, M. N. Myers, Anal. Chem., 46, 1917 (1974).
- J. C. Giddings, K. D. Caldwell, "Field-Flow Fractionation," in Physical Methods of Chemistry, B. W. Rossiter, J. F. Hamilton, eds., John Wiley & Sons, New York, 1989, Vol. 3B, Chap. 8, p. 867.
- M. Martin, P. S. Williams, "Theoretical Basis of Field-Flow Fractionation," in Theoretical Advancement in Chromatography and Related Separation Techniques, F. Dondi, G. Guiochon, eds., NATO ASI series C: Mathematical and Physical Science, Kluwer, Dordrecht, 1992, Vol. 383, p. 513.
- D. Melucci, G. Gianni, G. Torsi, A. Zattoni, P. Reschiglian, J. Liq. Chromatogr. & Rel. Technol., 20, 2615 (1997)
- E. Urbankova, J. Chmelik, J. Liq. Chromatogr. & Rel. Technol., 20, 2637 (1997).
- 10. G. Karaiskakis, A. Koliadima, Chromatographia, 28, 31 (1989).
- 11. A. Koliadima, G. Karaiskakis, J. Chromatogr., 517, 345 (1990).
- 12. E. Ruckenstein, A. Marmur, W. N. Gill, J. Colloid Interface Sci., **61**, 183 (1977).
- 13. R. Hogg, T. W. Healy, D. W. Fuersteuan, Trans. Faraday Soc., **62**, 1638 (1966).
- P. C. Hiemenz, Principles of Colloid and Surface Chemistry, Marcel Dekker Inc., New York, 1977, p. 457.
- 15. A. Athanasopoulou, G. Karaiskakis, Chromatographia, 40, 734 (1995).

16. A. Koliadima, G. Karaiskakis, Chromatographia, 39, 74 (1994).

- 17. J. C. Giddings, G. Karaiskakis, K. D. Caldwell, M. N. Myers, J. Colloid Inter-face Sci., **92**, 66 (1983).
- 18. A. Koliadima, G. Karaiskakis, J. Liq. Chromatogr., 11, 2863 (1988).
- G. Karaiskakis, M. N. Myers, K. D. Caldwell, J. C. Giddings, Anal. Chem., 53, 1314 (1981).
- 20. F-S. Yang, K. D. Caldwell, J. C. Giddings, J. Colloid Interface Sci., 92, 81 (1983).

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